

Average refractive index of tendon as a function of water content

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Abstract. Experimental data on the dependence of the average refractive index of rat tail tendon (RTT) on water content are reported. Using optical coherence tomography, the average group refractive index (at a wavelength of 930 nm) and cross-section area of rat tail tendon fascicle specimens during their air-drying and rehydration were monitored. The dependence of the average group refractive index of RTT (n_g) on the volume fraction of water (C_w) has been found to be nonlinear and to be well approximated by the quadratic polynomial $n_g = 1.5713 - 0.1969C_w - 0.0328(C_w)^2$. The reported data are shown to be in good agreement with previously published data for bovine cornea. © 2018 Journal of Biomedical Photonics & Engineering.

Keywords: collagen; tissue; refractive index; water content; hydration.

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1 Introduction

One of the optical parameters that can be directly measured for biological tissues is the average refractive index of the tissue [1-15]. In the literature, one can find estimates of the average refractive index for many types of tissues (cornea, sclera, dermis, epidermis, brain tissue *etc.*; see, *e.g.*, [1, 2, 4, 7-13, 15, 16]). The average refractive index depends on tissue composition and, in particular, on the water content [1]. The water content is one of the most variable and important tissue parameters, and the possibility to evaluate it from the measured values of the average tissue refractive index is of great practical interest. However, to date direct measurements of the average tissue refractive index as a function of tissue water content have been made only for a small number of tissues, *e.g.*, muscle tissue [13, 15] and brain tissues [1, 16]. Of collagenous tissues, as far as we are aware, reliable estimates of the average refractive index as a function of water content were obtained only for cornea [7], which is a tissue with a very high water content – normal cornea contains about 76 wt % water and 15 wt % collagen [17]. In this paper we present experimental estimates for the average tissue refractive index as a function of water content for tendon, a collagenous tissue with the highest normal collagen content—tendons contain typically 24–38 wt % collagen and 55–72 wt % water [18-20]. The experiments reported here were performed on rat tail tendon (RTT) fascicles. Fascicles are secondary collagen bundles of tendons [20-22]. Their composition and collagen fibril packing are similar to those of collagen fibers of dermis and sclera [17]. But fascicles, whose diameter typically lies within the range from 250 to 500 μm , are tens times thicker than dermal and scleral collagen fibers, which makes them quite convenient for experimentation. Due to their relatively simple structure, ease of extraction and manipulation, and ready availability, RTT fascicles are a very popular model object for studying physical and physiological properties of collagen fibers and collagenous tissues. For example, they are used in studying the effect of immersion agents, which are employed in the immersion optical clearing technique [23], on collagen bundles (see, *e.g.*, [24]). The quantitative data presented here can be used in such studies as reference data. For example, these data can be used for evaluating the duration of the solely dehydration stage of the immersion agent – biotissue interaction.

2 Methods

In order to determine the average refractive index of RTT fascicles as a function of water content, we monitored, using optical coherence tomography (OCT), the cross-section area and average refractive index of RTT fascicle specimens during their air-drying from the native state to the air-dry state and in the process of rehydration of dried specimens in normal saline solution (NSS: aqueous solution of 0.9 wt % NaCl). A series of measurements began with the measurement of the cross-

section area and refractive index of the sample in its initial (native) state. The fascicle, being submerged in NSS, was mounted, in a slightly stretched state, on an object-plate using binder clips and covered with a cover-slip. Then it was placed in the OCT beam path so that the direction of the fascicle was perpendicular to the B-scan direction in order to obtain an OCT-scan of the cross-section of the fascicle (Fig. 1a). After that, the cover-slip was removed, and the NSS was quickly removed from the surface of the object-plate with a filter paper. The (non-covered) sample was placed on the OCT scanner stage in the same position as before in order to monitor the sample parameters during its dehydration in air at room temperature (Fig. 1b, c). To estimate the parameters of the sample in the standard air-dry state, the sample was allowed to dry for 1 hr at 105°C (Fig. 1d). For rehydration measurements, the dried fascicle on the object-plate was surrounded by a large amount of NSS and covered by a cover-slip.

One of the parameters that was traced in the experiment was the coefficient of fascicle volume change (volume factor)

$$k_s = V/V_0, \quad (1)$$

where V is the current value of tissue volume and V_0 is the volume of the tissue in the initial state. In our experiments, the tendon fascicle with fixed ends remained slightly stretched during both the dehydration and following rehydration, *i.e.* the length of the fascicle did not change. Therefore the volume factor can be calculated as

$$k_s = S/S_0, \quad (2)$$

where S and S_0 are the values of the cross-section area at the moment of measuring and in the initial state, respectively.

The measured values of k_s were used for estimating the water volume fraction C_w in the tissue. By definition,

$$C_w = V_w/V, \quad (3)$$

where V is the volume of the tissue and V_w is the volume of water in the tissue. On the assumption that to a good accuracy

$$V = V_{dry} + V_w, \quad (4)$$

where V_{dry} is the volume of the tissue in the dry state, it follows from (1) and (3) that

$$C_w \approx \frac{k_s - k_{s,dry}}{k_s}, \quad (5)$$

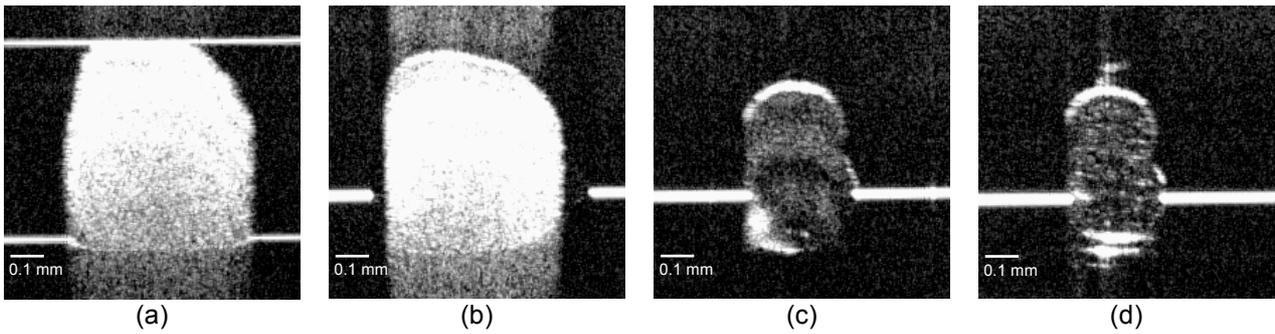


Fig. 1 OCT-images of a transverse cross-section of an RTT fascicle (a) in normal saline solution (native state), (b) after air-drying at room temperature for 1.7 min, (c) after air-drying at room temperature for 24 min, and (d) after air-drying at 105°C for 1 hour.

where k_{sdry} is the value of k_s for the dry state. For the native state, $C_w = 1 - k_{sdry}$.

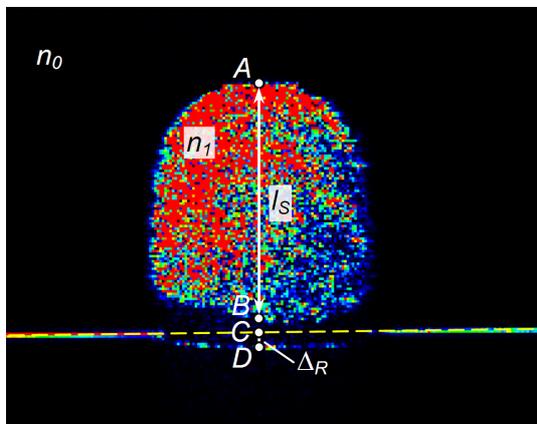


Fig. 2 Parameters used in the calculations of the average group refractive index of the tissue from an OCT-image of the specimen.

To estimate the average refractive index of the tissue using OCT, we employed a common method [3, 7, 25, 26] in which the group refractive index and physical thickness of the sample are determined from the measured values of the optical path length $l_s = n_1 d$, where d is the physical thickness of the sample for the given A-scan line and n_1 is the average group refractive index of the sample for this scan line, and shift Δ_R of the image of a reflecting surface arranged beyond the sample with respect to its position in the absence of the sample (in our experimental geometry, as in [26], the role of this reflecting surface is played by the frontal surface of the object–plate (glass substrate); see Fig.2). In an example in Fig. 2, where an OCT-image of a transverse section of an RTT fascicle is shown, apart from a common scale factor, $l_s = |AB|$ and $\Delta_R = |CD|$. Given l_s and Δ_R , the average refractive index of the sample n_1 for the A-scan line under consideration can be calculated as

$$n_1 = \frac{n_0 l_s}{l_s - \Delta_R},$$

where n_0 is the group refractive index of the surrounding medium (in our experiments, air or NSS). We calculated the average group refractive index of the specimen for the given cross-section as an average over several (3 to 5) A-scan lines. The group refractive index of the surrounding medium was calculated from dispersion data for the phase refractive index of the medium using the following common relation [27, 28]:

$$n_g(\lambda_0) = n_p(\lambda_0) - \lambda_0 \left. \frac{dn_p}{d\lambda} \right|_{\lambda=\lambda_0}, \quad (6)$$

where n_p is the phase refractive index, n_g is the group refractive index, and λ_0 is the central wavelength of the probing radiation in vacuum. For the OCT-system that we used, ThorLabs-OCP930SR, $\lambda_0 = 930$ nm. We took $n_g(930 \text{ nm}) = 1$ for air and $n_g(930 \text{ nm}) = 1.3416$ for NSS. The phase refractive index of NSS was assumed to be approximately equal to the phase refractive index of water. For water, we used the dispersion data reported in [29], which gave the mentioned value of 1.3416.

The area S [see(2)] was calculated using the formula

$$S = \frac{S_{\text{OCT-pix}} k_{pm^2}}{n_{gt}}, \quad (7)$$

where $S_{\text{OCT-pix}}$ is the area of the specimen cross-section in the OCT-image in pixels, k_{pm^2} is an instrument scale factor (in our case, $k_{pm^2} = 1.2632 \cdot 10^{-5} \text{ mm}^2/\text{pixel}$), and n_{gt} is the value of the average group refractive index of the specimen calculated from the OCT data for this cross-section as described above. To find $S_{\text{OCT-pix}}$ we used the open-source image analysis software Icy [30].

An example of the measured time-dependences of the volume factor k_s (2) and the average group refractive index of the tissue in the process of air-drying for a sample of RTT fascicle is presented in Fig. 3. It is clearly seen that the shrinkage of the tissue due to water

loss is accompanied by an increase in its average refractive index.

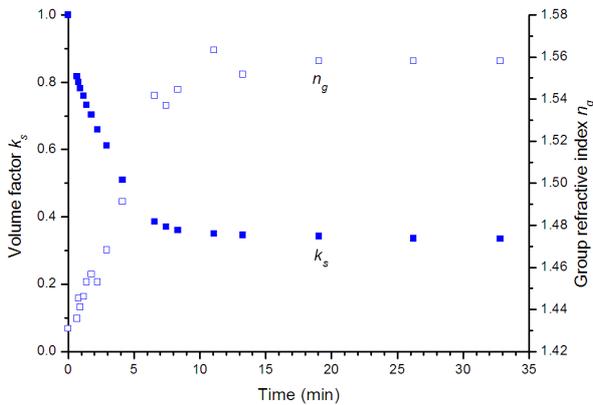


Fig. 3 Change of the volume factor k_s and average refractive index n_g of an RTT fascicle specimen during air-drying at room temperature.

Since the samples were relatively small in size, the boundaries of the sample and the upper boundary of the object-plate under the sample were clearly visible in OCT images even when the sample was in the native state, and their position could be determined with good accuracy. This ensured high accuracy of cross-section area measurements, which is confirmed by a small scatter of points in k_s vs time plots (as an example see Fig. 3). At the same time the samples were sufficiently thick for the relative accuracy of group refractive index measurements to be of the order of 0.3–0.5%.

3 Samples

Fascicles were excised from the tails of mature rats within one hour after decapitation, then immediately immersed in NSS. All refractive-index measurements were performed on fascicles ranging in diameter from 300 to 400 μm . The storage time of samples in NSS before they were used in experiments did not exceed 7 days. No statistically significant change (Friedman test, probability value $p = 0.32$) in the average refractive index of RTT fascicle specimens was observed during 7-day storage in NSS (see Fig. 4).

The studies were approved by the Ethics Committee of Saratov State Medical University (Saratov, Russia).

4 Results and discussion

Fig. 5 shows the measured dependences of the average refractive index, n_g , on the volume factor k_s for four samples of RTT fascicles, samples 1, 2, 3, and 4. The OCT data for sample 1 were collected during 30 min when this sample was dried at room temperature. Sample 2 was dried at room temperature for 2 hr, then held for 1 hr at 50°C, and finally held for 1 hr at 105°C. Sample 3 was dried for 40 min at room temperature and then for 1 hr at 105°C. Sample 4 was first dried at room temperature for 20 min and then placed in NSS for

rehydration. From Fig. 5, it can be seen that the data for different samples are in good agreement, as are the data for drying and rehydration of sample 4.

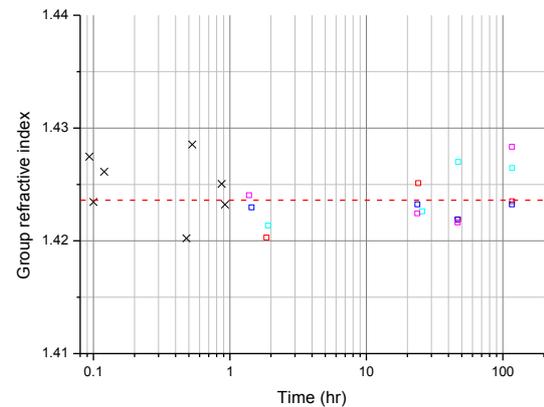


Fig. 4 Time and intersample variation of the measured refractive index for RTT specimens stored in normal saline solution (NSS). Abscissa: time of sample storage in NSS. The squares of the same color correspond to the same RTT fascicle specimen. The crosses correspond to different specimens. The red dash line shows the average value of n_g over all experimental points on this graph.

An average value of n_g for RTT fascicles in the native state was found to be 1.423 ± 0.003 (mean \pm SD; 12 samples). This value is shown by the black triangle in Fig. 5. An average value of $k_{s,dry}$ was 0.323 ± 0.003 (mean \pm one-half of the range; 3 samples), and, consequently, the average volume fraction of water for fascicles in the native state can be estimated as 0.677 ± 0.003 [see (5)]. Taking 0.677 ± 0.003 for C_w and 1.34 for the specific gravity ρ_{dry} of the dry tissue [31], we may estimate the average weight water content for the samples in the native state as 61 ± 0.3 wt %.

In Fig. 6, the measured values of n_g for RTT are plotted against the volume water fraction C_w , C_w being calculated by (5) at $k_{s,dry} = 0.323$. Kim *et al.* [7] reported the measured values of the group refractive index as a function of a hydration parameter H for bovine cornea at $\lambda_0 = 819.9$ nm. The hydration H of tissue is defined as the ratio of the weight of water to the dry weight. Having calculated C_w from H as

$$C_w = \frac{H\rho_{dry}}{1 + H\rho_{dry}} \quad (8)$$

at $\rho_{dry} = 1.34$, we transferred the experimental data for cornea [7] to Fig. 6 for comparison. It can be seen from the figure that the data for tendon and cornea fit rather well.

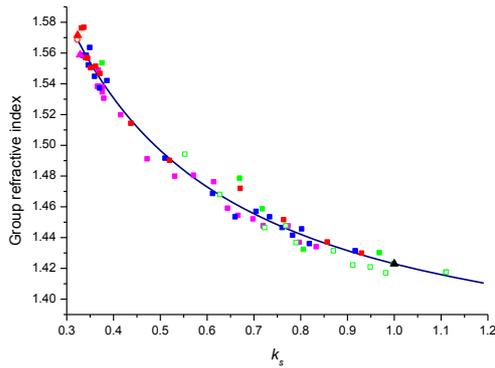


Fig. 5 Group refractive index of RTT fascicles vs volume factor k_s . The blue squares (■) show experimental points (k_s, n_g) for sample 1 (air-dried at room temperature). The red symbols (■, ▲, ○) represent the data for sample 2: the squares (■) are the points obtained during air-drying at room temperature; the open circle (○) and solid triangle (▲) show the points obtained on air-drying at 50°C and 105°C, respectively. Purple symbols (■, ▲) show the data for sample 3: the squares (■) are for air-drying at room temperature, and the triangle (▲) is for air-drying at 105°C. The green symbols (■, □) are for sample 4: the solid squares (■) are for the stage of air-drying, and open squares (□) are for the stage of rehydration (80 min). The black triangle (▲) shows the average value of n_g of RTT fascicles in the native state (over 12 specimens).

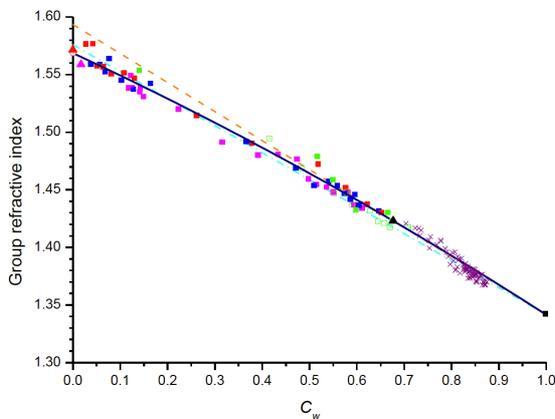


Fig. 6 Group refractive index of RTT fascicles vs volume water content C_w . The symbols are the same as in Figure 5. The crosses (×) represent the experimental points for bovine cornea [7] ($\lambda_0 = 819.9$ nm). The solid line (—) and cyan dash-dot (— · —) line represent respectively the approximating quadratic (Eq.9) and linear (Eq.10 with $n_{dry} = 1.576$) polynomials to the data for RTT. The orange dash line (— — —) is the straight line (Eq.10 with $n_{dry} = 1.594$) passing through the points (C_w, n_g)=(1, 1.3416) (water) and (0.677, 1.423) (the average for the native state of RTT).

The experimental data for RTT are well approximated by the quadratic polynomial

$$n_g = 1.5713 - 0.1969C_w - 0.0328C_w^2 \quad (9)$$

(Fig. 6). The coefficients of this expression were found by the least squares method under the additional conditions that $n_g(C_w)$ must be equal to the group refractive index of water $n_w = 1.3416$ at $C_w = 1$ and to the reliable estimate $n_g = 1.423$ at $C_w = 0.677$ (native state). At $C_w = 0$, approximation (9) gives $n_g = 1.5713$, which is close to the measured values of the group refractive index for the dry state, n_{dry} . We estimated the accuracy of this and subsequent approximations using the parameter $\sigma_{fit} = \sqrt{r_{ss}/n}$, where r_{ss} is the residual sum of squares and n is the number of experimental points. For approximation (9), $\sigma_{fit} \approx 5.4 \cdot 10^{-3}$. When approximating the experimental data by a linear function

$$n_g = (1 - C_w)n_{dry} + n_w C_w, \quad (10)$$

which corresponds to the Gladstone-Dale law [23, 31, 32], with given $n_w = 1.3416$, σ_{fit} is minimum (about $6.7 \cdot 10^{-3}$) when $n_{dry} = 1.576$ (Fig. 6, cyan line). This approximation gives a significantly underestimated value of n_g (1.417) for the native state. The approximation of the RTT data by the straight line passing through the points (C_w, n_g) = (1, 1.3416) (water) and (0.677, 1.423) (native state), the red line in Fig. 6, ensures a rather good accuracy for the range $0.52 \leq C_w \leq 1$ ($\sigma_{fit} \approx 4 \cdot 10^{-3}$) but gives a significantly overestimated value for n_{dry} (1.594). The significant deviation of the actual $n_g(C_w)$ curve from this approximating line in the region $C_w \leq 0.52$ may be attributed to a change of the mode of hydration. In the literature, two modes of collagenous tissue hydration are distinguished [33-36]. For relatively high values of H ($H > H_c$, where H_c is a critical value, the so-called fibrillar saturation point [35]), changes in the total water content in the tissue lead to changes of the water content in extrafibrillar space while the water content in collagen fibrils remains unchanged [7, 33-36]. In the alternative mode ($H < H_c$), changes in the total water content are accompanied by changes in water content both in collagen fibrils and extrafibrillar space [33-36]. For RTT, $H_c \approx 0.82$ [32, 35], which corresponds to $C_w \approx 0.52$.

Fig. 7 shows the dependence of the phase refractive index n_p of RTT on C_w . The phase refractive index was calculated using approximating function (9) and Eq. (6) on the assumption that

$$\left. \frac{dn_p}{d\lambda} \right|_{\lambda=\lambda_0} = (1 - C_w) \left. \frac{dn_{dry}}{d\lambda} \right|_{\lambda=\lambda_0} + C_w \left. \frac{dn_w}{d\lambda} \right|_{\lambda=\lambda_0}.$$

The values of $dn_{dry}/d\lambda$ and $dn_w/d\lambda$ were calculated from the dispersion data for gelatin [37] and water [29], respectively. We did not find in the literature any reliable experimental data on the wavelength dependence of the phase refractive index of dry collagen in the spectral range under consideration. Gelatin is known to be produced by partial hydrolysis of animal collagen and to have a very similar chemical structure to collagen. For this reason, we used the available data for dry gelatin. For the phase refractive index of gelatin we used the dispersion formula $n = 1.53 + 1788/\lambda_{nm}^2 + 5.73 \cdot 10^8/\lambda_{nm}^4$, where λ_{nm} is the numerical value of the wavelength λ in nanometers [37].

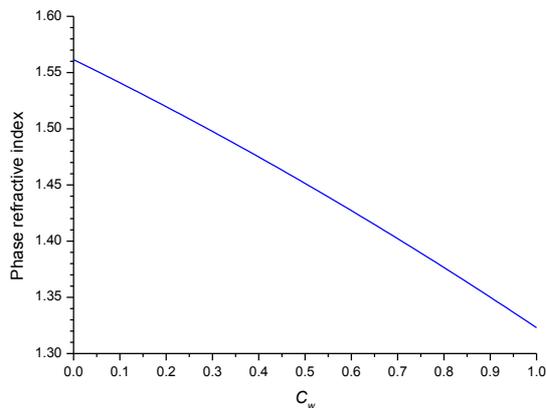


Fig. 7 Tentative estimate of the average phase refractive index of RTT fascicles as a function of volume water content.

5 Conclusion

The average group refractive index of RTT fascicles has been measured as a function of water content using OCT. For the native state, this refractive index has been found to be 1.423 ± 0.003 at a wavelength of 930 nm. In the range $0.52 \leq C_w \leq 1$, the average group refractive index of RTT fascicles has been found to change almost linearly with C_w . Over the entire range of C_w values, $0 \leq C_w \leq 1$, the experimental data are well fitted by the quadratic polynomial (9).

Disclosures

The authors declare that there are no conflicts of interest related to this article.

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